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MORPHOMETRIC ANALYSIS OF FIVE SPECIES OF PONYFISHES (FAMILY:LEIOGNATHIDAE) COLLECTED FROM TAMILNADU COAST, INDIA

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AUTHORS' CONTRIBUTIONS

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

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Original Research Article

ABSTRACT

Leiognathids, commonly known as ponyfishes are characterised by their protrusible mouth and there are a lot of controversies in their nomenclature. In this study, a total of five species of family Leiognathidae were collected from the Parangipettai landing centre, Tamilnadu, India. For morphometric analysis, 24 morphometric data points were collected for each species and their variation was studied. There were significant variations among the species, with minor overlap. Out of 24 characters, 14 were unique to each species. The features on pectoral and pelvic fins, upper and lower jaw lengths and pre-orbital length were highly significant variable characteristics among the ponyfish studied. Among the five species, the within-species variation was more in *Gazza minuta* and *Eubleekeria splendens*. But the within -species variation was less in *Karalla dussumieri* and *Nuchequula gerreoides*.

Keywords: Ponyfish; leiognathidae; morphometric; principal component analysis.

1. INTRODUCTION

Morphometric and meristic characters have long been used to separate species, populations and races. It also helps identify species and find sexual dimorphism [1]. Morphometric studies are often used to support descriptions of new species based on morphological differences in closely related species [2,3,4]. Some studies emphasise new taxonomic descriptions by integrating new analysis tools into morphometry [5,6]. Morphological analysis can also be used for phylogenetic assessment [7], phenotypic plasticity [8, 9] and to study various fish conditions [10].

Ponyfishes, also called as silverbellies, belong to the family Leiognathidae and are one of the most

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commercially important fish groups in fishing industry due to the high quantity of bone and fatless flesh that provide plenty of calcium and protein [11]. Ponyfishes have bacterial light organs in the throat region from which light is spread through the belly [12]. Leiognathidae was initially composed of only two genera, Leiognathus and Gazza. Starting from the evolution or retrieval of the genus Secutor, the family currently includes at least nine genera [13, 14]. Many of the species are synonyms and this leads to lot of controversy in the nomenclature. Hence, the family Leiognathidae is in need of taxonomic revision [15, 16]. Morphometric data is commonly analysed using univariate statistics (ANOVA) and multivariate statistics, such as principal components analysis (PCA). PCA is always superior to univariate analysis because it considers the covariance or correlation structure of the data and considers relationships among all measured morphometric and shape characters [17].

In India, the morphometric and meristic character analysis in Leiognathidae is very rare and no previous work is available to distinguish the species. To fill the paucity, the study was planned to delineate five species belonging to the Parangipettai region of the Tamilnadu coast, using morphometric analysis.

2. MATERIALS AND METHODS

2.1 Sample Collection

A total of 450 fish were collected from the Parangipettai landing centre (11.4831° N, 79.7729° E) from January to June, 2019. After collection, the samples were preserved in ice and brought to the laboratory for identification and morphometric measurements. The species were identified by the FAO sheet [18].

2.2 Morphometric Measurements and Data Analysis

Twenty four morphometric characters were measured using a digital Vernier caliper with an accuracy of 0.01mm. Here, most of the major measurements were converted as percentage of standard length rather than total length or fork length, as per the previous study by Onsoy et al. [19]. An ANOVA was carried out for all morphometric characters and *P*-values were calculated. PCA was performed using SPSS 15.0 software. Characters that were used in the present study and their abbreviations are listed in Table 1. The diagrammatic representations of the measured characters are displayed in Fig. 1 and Fig. 2.

Table 1. Morphometric characters and their abbreviations used in this study

Sl. No.	Characters	Abbreviation
1.	Standard Length	SL
2.	Head length	HL
3.	Pre-dorsal Fin Length	PDL
4.	Pre-pectoral Fin Length	PPcL
5.	Pre-anal Fin Length	PAL
6.	Pre-ventral Fin Length	PVL
7.	Pectoral Fin Length	PcFL
8.	Pelvic Fin Length	PIFL
9.	Anal Fin base Length	ABL
10.	Dorsal Fin base Length	DBL
11.	Caudal Peduncle Length	CPL
12.	Caudal Peduncle Width	CPW
13.	Head Width	HW
14.	Body Width	BW
15.	1 st Dorsal Fin Spine Length	1 st DSL
16.	2 nd Dorsal Fin Spine Length	2 nd DSL
17.	3 rd Dorsal Fin Spine Length	3 rd DSL
18.	1 st Anal Fin Spine Length	1 st ASL
19.	2 nd Anal Fin Spine Length	2 nd ASL
20.	3 rd Anal Fin Spine Length	3 rd ASL
21.	Eye diameter	ED
22.	Pre-orbital Length	POL
23.	Upper Jaw Length	UJL
24.	Lower Jaw Length	LJL

Characters	Eubleekeria	Karalla	Nuchequula	Gazza minuta	Equulites
	splendens	dussumieri	gerreoides		leuciscus
SD (mm)	76.80 ± 8.50	57.67±5.29	59.66±5.37	84.97± 8.43	84.97±13.05
HL (mm)	24.76±2.98	18.98 ± 1.77	19.43±1.35	26.99 ± 2.40	22.53±3.34
HL (% of SL)	32.23±1.06 ^a	32.93 ± 1.29^{b}	32.64 ± 1.62^{a}	31.82 ± 1.34^{a}	$26.55 \pm 0.74^{\circ}$
PDL (% of SL)	38.03±2.64 ^a	39.21 ± 1.96^{b}	$42.01 \pm 1.53^{\circ}$	$41.57 \pm 1.62^{\circ}$	38.50 ± 1.02^{a}
PPcL (% of SL)	36.19±1.31 ^a	36.96±1.81 ^b	$35.61 \pm 2.78^{a,c}$	$34.92 \pm 1.46^{\circ}$	31.19±1.15 ^d
PAL (% of SL)	53.05±2.01 ^a	52.99 ± 2.10^{a}	53.06 ± 2.22^{a}	$52.93{\pm}2.18^{\mathrm{a}}$	50.97 ± 2.23^{b}
PVL (% of SL)	35.08 ± 1.55^{a}	36.42 ± 1.94^{b}	36.49 ± 1.49^{b}	$33.13 \pm 2.07^{\circ}$	32.40 ± 1.41^{d}
PcFL (% of SL)	25.31±1.31 ^a	22.39±1.04 ^b	$21.48 \pm 1.15^{\circ}$	19.09 ± 1.15^{d}	17.66±0.75 ^e
PIFL (% of SL)	16.47 ± 0.88^{a}	17.21 ± 0.94^{b}	$14.49 \pm 0.85^{\circ}$	14.01 ± 1.09^{d}	12.02±0.77 ^e
ABL (% of SL)	41.82 ± 1.51^{a}	40.13 ± 1.83^{b}	40.89 ± 2.32^{b}	39.96 ± 1.38^{b}	40.47 ± 1.28^{b}
DBL (% of SL)	55.48 ± 1.71^{a}	51.73 ± 1.93^{b}	52.56 ± 1.64 ^c	48.84 ± 1.69^{b}	51.68 ± 1.37^{d}
CPL (% of SL)	5.33 ± 0.47^{a}	4.93 ± 0.57^{b}	$5.76 \pm 0.75^{\circ}$	$5.22 \pm 0.54^{\mathrm{a}}$	4.89 ± 0.42^{b}
CPW (% of SL)	8.14 ± 0.45^{a}	7.26 ± 0.44^{b}	$6.52 \pm 0.67^{\circ}$	$6.35 \pm 0.34^{c,d}$	6.28 ± 0.37^{d}
HW (% of SL)	30.01±1.25 ^a	27.62 ± 1.02^{b}	28.31 ± 2.56^{b}	$30.47 \pm 1.34^{\circ}$	23.39±0.83 ^d
BW (% of SL)	55.15 ± 2.06^{a}	49.21 ± 1.58^{b}	$45.39 \pm 1.13^{\circ}$	$45.22 \pm 1.63^{\circ}$	41.40 ± 1.67^{d}
1 st DSL (% of SL)	5.42 ± 0.55^{a}	4.58 ± 0.54^{b}	$3.25 \pm 0.35^{\circ}$	$2.52 \pm 3.46^{c,d}$	2.18 ± 0.26^{d}
2 nd DSL (% of SL)	$22.14{\pm}1.70^{a}$	25.58 ± 2.82^{b}	22.09±0.35 ^a	18.83±1.33 ^c	38.85 ± 2.38^{d}
3 rd DSL (% of SL)	19.55 ± 1.54^{a}	21.59 ± 1.44^{b}	$18.01 \pm 1.42^{\circ}$	16.75 ± 1.24^{d}	20.42 ± 1.66^{e}
1 st ASL (% of SL)	6.12 ± 0.68^{a}	5.89 ± 0.52^{a}	$4.94{\pm}0.49^{ m b}$	$2.81\pm0.32^{\circ}$	2.17 ± 0.36^{d}
2 nd ASL (% of SL)	18.92±1.51 ^a	18.54 ± 1.17^{a}	17.83 ± 1.21^{b}	$15.41 \pm 0.87^{\circ}$	16.35 ± 1.75^{d}
3rd ASL (% of SL)	14.87 ± 1.19^{a}	14.43 ± 1.01^{b}	13.68±1.25 ^c	12.52 ± 0.90^{d}	11.59±0.89 ^e
ED (% of HL)	37.66±1.71 ^a	33.49 ± 2.19^{b}	32.97 ± 1.97^{b}	$36.15 \pm 1.54^{\circ}$	30.60 ± 1.70^{d}
POL (% of HL)	19.56 ± 1.81^{a}	33.49 ± 1.97^{b}	26.52±2.27 ^c	25.08 ± 1.83^{d}	23.75±1.95 ^e
UJL (% of HL)	45.96 ± 2.58^{a}	54.97 ± 2.49^{b}	$60.87 \pm 2.10^{\circ}$	68.73 ± 2.81^{d}	59.34±2.78 ^e
LJL (% of HL)	40.50 ± 2.14^{a}	45.02 ± 2.41^{b}	$50.34 \pm 2.27^{\circ}$	60.34 ± 2.76^{d}	46.67±2.19 ^e

Table 2. Morphometric characters in five ponyfishes

Values on the raw sharing the common superscript are not significantly varied (P < 0.05)

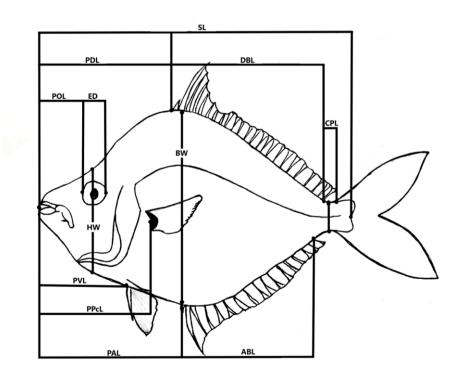


Fig. 1. Diagrammatic representation of morphometric characters used in this study

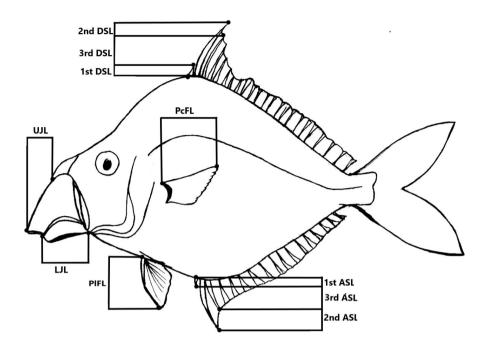


Fig. 2. Diagrammatic representation of morphometric characters used in this study

3. RESULTS AND DISCUSSION

The specimens collected were identified as *Eubleekeria splendens* (Cuvier, 1829), *Karalla dussumieri* (Valenciennes, 1835), *Nuchequula gerreoides* (Bleeker, 1851), *Gazza minuta* (Bloch, 1795) and *Equulites leuciscus* (Gunther, 1860). Average values as percentage of SL or HL for each character are given in Table 2.

Among the 24 morphological values, most of them showed high significant variation at the level of P <0.05, in five species. In the case of HL, a significant variation was observed between all species except E. splendens (38.03±2.64), G. minuta (31.82±1.34) and N. gerreoides (32.64±1.62) and the lowest value of HL was observed in E. leuciscus (26.55±0.74). Variation in PDL was significant among all species except E. splendens (38.03±2.64) and E. leuciscus (38.50±1.02). PPcL also showed a significant variation among the three species, except for E. splendens (36.19±1.31) and Ν. gerreoides (35.61±2.78). However, for PAL, only E. leuciscus (50.97±2.23) showed a significantly lower value, and it was almost similar in other species. PVL values were similar in K. dussumieri (36.42±1.94) and N. gerreoides (36.49±1.49) but varied in other species. The ABL showed significant variation only in E. splendens (41.82±1.51). DBL was significantly similar in K. dussumieri (51.73±1.93) and G. minuta (48.84 \pm 1.69). CPL was significantly similar in E. splendens and G. minuta and also in G. minuta and E. leuciscus. The HW value was highly variable except

in K. dussumieri (27.62±1.02) and N. gerreoides (28.31 \pm 2.56). Like HW, the BW value also showed a significant variation, except for G. minuta (45.22 ± 1.63), and N. gerreoides (45.39 ± 1.13). The 1st DSL value showed significant variation in E. splendens and K. dussumieri. But the value of G. minuta was similar to that of N. gerreoides and E. leuciscus. The second DSL showed the most significant variation of all the characters among the species. 1st and 2nd ASL values showed significant variation in all species except E. splendens and K. dussumieri. PVL, DBL, HW, BW, 1st ASL, 2nd ASL, and ED showed a high level of variation, but the variation was insignificant only for K. dussumieri (14.43 ± 1.01) and N. gerreoides (32.97±1.97). Among all the morphometric characters, PcFL, PIFL, 3rd DSL, 3rd ASL, POL, UJL, and LJL showed high significant variation between all the five species.

In Eigen values of PCA, PC1 showed a 48.8% confidence level and PC2 showed a 30.2% one. Variations in PC1 were mainly due to HL, PDL, PPcL, PVL, PcFL, PIFL, ABL, DBL, CPW, BW, 1st DSL, 3rd DSL, 1st, 2nd and 3rd ASL, ED, POL, UJL, and LJL. Among these, the highest variation was shown by UJL and LJL. Variations along PC2 were due to HL, PDL, PPcL, PAL, PVL, PcFL, PIFL, CPL, CPW, HW, BW, 1st, 2nd and 3rd DSL, 1st ASL, 3rd ASL, and LJL. All the characters that were selected for the present study showed considerable weightage that can be used to discriminate at species level, in either PC1 or PC2, as shown in Table 3.

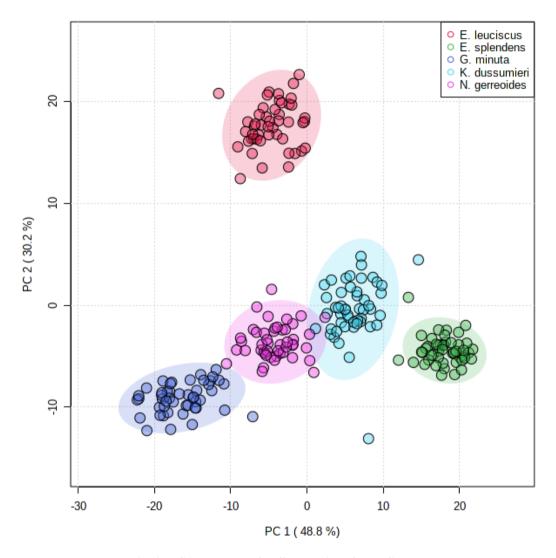


Fig. 3. PCA score plot for five species of ponyfishes

The PCA scatter plot is given in Fig. 3 and it is evident that each group was separately clustered with minor overlap. In this plot, G. minuta and N. gerreoides showed a negative correlation, while K. dussumieri and E. splendens showed a positive correlation along PC1. Except for E. splendens, all of the species had a negative correlation along PC2, while only E. leuciscus had a high positive correlation along PC2 and was clustered far away from the other species. Along with PC1, more correlation was exhibited between N. gerreoides and K. dussumieri, followed by N. gerreoides and G. minuta, and then by K. dussumieri and E. splendens. The least correlation along PC1 was observed between G. minuta and E. splendens. When considering PC2, E. leuciscus showed high variation from all other species.

Morphometric variation among five ponyfish from five genera was analysed in the present study. Morphological variation increases from lower to higher taxa, and it is mainly due to genetic variation [20]. Though species constitute the smallest taxonomic group, morphological variations are evident in populations also [21]. Morphological variation in fish can affect their diet and can modify their feeding behavior and feeding patterns [22,23]. Even though morphological studies have various applications, the present study is mainly focused on discriminating the ponyfish, which are in turn used in fisheries management as in earlier studies [24].

The morphological data of *E. splendens* are consistent with the previous findings [25,26]. In *E. splendens*, the values of PVL, PcFL, PIFL, ABL, DBL, CPW, HW, BW, 1st DSL, 3rd DSL, 3rd ASL, ED, POL, UJL, and LJL showed significant variations from all other species. Among the five species, *E. splendens* showed maximum similarity to *N. gerreoides* and maximum variation to *E. leuciscus*. These findings are consistent with previous findings by Seth et al. [24], who found

that *N. gerreoides* had a significant shape difference from the species *E. leuciscus* based on Procrustes distance studies. Morphometric characters of *K. dussumieri* are similar to earlier reports by Abraham et al. [25]. *Karalla dussumieri* showed the least variation compared to *N. gerreoides* with five morphometric characters (PAL, PVL, ABL, HW, and ED). *Karalla dussumieri* can be distinguished from *E. leuciscus* by most of the characters, and this was confirmed by molecular data in the previous report [27].

Table 3. Morphometric variable loadings for the
first and second principal components in five
ponyfishes

Characters	PC 1 (48.8%)	PC 2 (30.2%)
HL	0.2779	0.7878
PDL	-0.4567	0.3673
PPcL	0.3544	0.6401
PAL	0.1033	0.3877
PVL	0.3473	0.3600
PcFL	0.7545	0.4540
PIFL	0.5889	0.5208
ABL	0.3032	0.0417
DBL	0.7224	-0.0352
CPL	0.0158	0.2635
CPW	0.7621	0.2627
HW	0.0953	0.8507
BW	0.7788	0.5091
1 st DSL	0.5457	0.2585
2 nd DSL	-0.0009	-0.9786
3 rd DSL	0.4692	-0.4542
1^{st} ASL	0.7462	0.4449
2^{nd} ASL	0.6581	0.1022
3 rd ASL	0.5969	0.4319
ED	0.2499	0.6495
POL	-0.6875	0.0265
UJL	-0.9746	0.0810
LJL	-0.9116	0.3438

For N. gerreoides, the morphometric characters of the present study are similar to those reported for N. mannusella by Chakrabarty and Sparks [16], but values of PVL, HW, UJL, and LJL are slightly higher in N. gerreoides. Values of PDL, PcFL, PIFL, DBL, CPL, CPW, 3rd DSL, 3rd ASL, POL, UJL, and LJL in N. gerreoides showed significant variation from the other four species. The same observations were recorded in a previous study by Seth et al. [24]. A morphological study on G. minuta by Yoshitaka et al. [28] and Jawad et al. [29] produced similar results as in the present study. G. minuta's PcFL, PlFL, 3rd DSL, 3rd ASL, POL, UJL, and LJL were significantly different from the other four species. Data obtained for E. leuciscus showed a similar kind of value to that of Abraham et al. [25]. The values of PcFL, PIFL, 3rd DSL, 3rd ASL, POL, ULJ, and LJL were significantly different from the other four species. *Equulites leuciscus* showed maximum similarity with *G. minuta* based on only three characters (ABL, CPW, and 1st DSL). This result is not in accordance with the previous study by Seth *et al.* [24]. In this study, PcFL, PIFL, 3rd DSL, 3rd ASL, POL, UJL, LJL, PVL, DBL, HW, BW, 1st ASL, 2nd ASL, and ED showed significant variation among the five species. These findings are consistent with Echem's [30] findings in shape variation studies on Leiognathidae.

A PCA should be performed, as the N:P ratio must be greater than three for an effective PCA interpretation [31]. In the present study, the N:P ratio was around 18, which shows a high confidence level. In the PCA scatter plot, each species is clustered into distinct groups with minor overlap. This overlapping may be due to the overall similarity and shape of the animals [32]. Along with PC1, maximum variation was observed between *E. splendens* and *G. minuta*. Interspecific variation was significantly higher than intraspecific, and is reflected in Fig. 3. In the PCA scatter plot, the intra-specific variation was less in *E. splendens* and was higher in *K. dussumieri*, but the intra-specific variation was reported as less by Seth *et al.* [24] in *K. dussumieri*.

4. CONCLUSION

Morphometric analysis of ponyfish in this study revealed that each species was separately scattered and minor overlapping was observed due to the deviation in morphological characteristics within the group. Minor overlap may be due to certain genetic similarities and some common characteristics they share since they belong to the same family. For the effective identification of ponyfishes, molecular level identification is very much needed.

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COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Remya M, Sherly W. A study of morphometry and meristic counts of *Oxyurichthys tentacularis*, Gobiidae (Valenciennes, 1837) from Ashtamudi lake- Kollam, Kerala. International Journal of Fisheries and Aquaculture Sciences. 2018;8 (1):13-18.

- 2. Creech S. A multivariate morphometric investigation of Atherina boyeri Risso, 1810 and *Atherina presbyter* (Cuvier 1829) (Teleostei:Atherinidae):Morphometrics evidence in support of the two species. Journal of Fish Biology. 1992;41:341–353.
- 3. Devaere S, Adriaens D, Verraes W. *Channallabes sanghaensis* sp. n., a new anguilliform catfish from the Congo river basin, with some comments on other anguilliform clariids (Teleostei, Siluriformes). Belgian Journal of Zoology. 2007;137:17–26.
- 4. Welsh SA, Wood RM. *Crystallaria cincotta*, a new species of darter (Teleostei:Percidae) from the Elk river of the Ohio river drainage, West Virginia. Zootaxa. 2008;1680:62–68.
- Das MK, Nelson JS Revision of the percophid genus Bembrops (Actinopterygii:Perciformes). Bulletin of Marine Science. 1996;59:9–44.
- 6. Vreven EJ, G. G. Teugels GG. Redescription of *Mastacembelus liberiensis* Boulenger, 1898 and description of a new West African spinyeel (Synbranchiformes:Mastacembelidae) from the Konkoure river basin, Guinea. Journal of Fish Biology. 2005;67:332–369.
- Morrison CL, Lemarie DP, Wood RM, King TL. Phylogeographic analyses suggest multiple lineages of *Crystallaria asprella* (Percidae:Etheostominae). Conservation Genetics. 2006;7:129–147.
- 8. Gillespie GJ, Fox MG. Morphological and life history differentiation between littoral and pelagic forms of pumpkinseed. Journal of Fish Biology. 2003;62:1099–1115.
- 9. Hard JJ, Winans GA, Richardson JC. Phenotypic and genetic architecture of juvenile morphometry in Chinook salmon. Journal of Heredity. 1999;90:597–606.
- Smith CD, Higgins CL, Wilde GR, Strauss RE. Development of a morphological index of the nutritional status of juvenile largemouth bass. Transactions of the American Fisheries Society. 2005;134:120–125.
- 11. Jeyaseelan PMJ, Ramanathan N, Sundararaj V, Venkataramanujam K, Devaraj M. Manual of fish eggs and larvae from Asian mangrove waters. UNESCO Publishing. 1998;147pp.
- 12. Paxton JR, Esch WN. Encyclopedia of Fishes. Academic Press Inc., London. 1998;240pp.
- Larson HK, Williams RS, Hammer MP. An annotated checklist of the fishes of the Northern Territory, Australia. Zootaxa. 2013;3696(1):1-293.
- 14. Kottelat M. The fishes of the inland waters of Southeast Asia:a catalogue and core

bibliography of the fishes known to occur in freshwaters, mangroves and estuaries. The Raffles Bulletin of Zoology. 2013;27:1-663.

- 15. Jones G. Revision of the Australian species of the fish family Leiognathidae. Australian Journal of Marine and Freshwater Research. 1985;36:559–613.
- 16. Chakrabarty P, Sparks JS. Phylogeny and taxonomic revision of *Nuchequula* Whitley 1932 (Teleostei:Leiognathidae), with the description of a new species. American Museum Novitates. 2007;3588:1–28.
- Bookstein FL, Chernoff B, Elder RL, Humphries JM, Smith GR Strauss RE. Morphometrics in evolutionary biology, Academy of Natural Sciences, Philadelphia. 1985;277pp.
- Thomson JM. FAO species identification sheet for fishery purposes, Western Indian-ocean. Fishing area 51. Fisher W and Bianchi (Eds.). FAO. Rome. 1984;Volume 2.
- Onsoy B, Tarkan AS, Filiz H, Bilge G. Determination of the best length measurement of fish. North-Western Journal of Zoology. 2011;7:178-180.
- 20. Robinson BW, Wilson DS. Genetic variation and phenotypic plasticity in a trophically polymorphic population of pumpkinseed sunfish (*Lepomis gibbosus*). Evolutionary Ecology. 1996;10:631–652.
- 21. Hanif AM, Siddik MAB, Islam A. Multivariate morphometric variability in sardine *Amblygaster clupeoides* (Bleeker, 1849), from the Bay of Bengal coast, Bangladesh. The Journal of Basic and Applied Zoology. 2019;80(1):53.
- 22. Motta PJ. Functional morphology of the feeding apparatus of ten species of Pacific butterflyfishes (Perciformes, Chaetodontidae): and ecomorphological approach. Environmental Biology of Fishes. 1988;22:39–67.
- 23. Norton SF. Capture success and diet of cottid fishes:the role of predator morphology and attack kinematics. The Ecological Society of America. 1991;72:1807–1819.
- 24. Seth JK, Barik TK, Mishra SS. Geometric morphometric approach to understand the body shape variation in the pony fishes (Leiognathidae) of Odisha Coast, India. Iranian Journal of Ichthyology. 2019;6(3):208-217.
- 25. Abraham KJ, Joshi KK, Murty VSR. Reproductive biology of *Leiognathus splendens* (Cuvier) from Kochi, South-west coast of India. Indian Journal of Fisheries. 2011;58(3):23-31.

- Acharya KV, Naik SD. Morphological characters and morphometric relationship of pony fish, *Leiognathus splendens* (Cuvier, 1829) Off Ratnagiri coast, Maharashtra, India. International Journal of Sciences and Applied Research. 2015;2(7):115-125.
- Banu F, Remanadevi V, Shalini G, Thangaraj M. Molecular variation and phylogenic status of ponyfish (Perciformes:Leiognathidae) in Karaikal, South India. Notulae Scientia Biologicae. 2020;12(2):251-257.
- 28. Yoshitaka Y, Yutaka Y, Hiroshi T. First record of the Leiognathid fish, *Gazza minuta* from Japan. Japanese Journal of Ichthyology. 1984;31:327-330.
- 29. Jawad LAJ, Al-Mamry JM. New record of the toothpony, *Gazza minuta* (Osteichthyes: Leiognathidae) from the coast of Muscat city at the sea of Oman, Sultanate of Oman. Thalassia Salentina. 2013;35:3-9.
- TR. Use 30. Echem of fish geometric morphometric markers for characterizing shape variations of selected fishes:Family Leiognathidae in the marine waters of Zamboanga City, Western Mindanao. Philippines. Available:https://www.essaycompany.com/essa ys/biology/using-fish-geometricmorphometric-2650.2018
- Kocovsky PM, Adams JV, Bronte CR. The effect of sample size on the stability of principal components analysis of truss- based fish morphometrics. American Fisheries Society. 2011;138:487-496.
- Echem RT. Geometric morphometric analysis of shape variation of *Sardinella lemuru*. International Journal of Advanced Research in Biological Sciences. 2016;3(9):91-97.

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